

SSV docs

Structural signatures



Manually curated database



Python scripts



Running online

Structural Signature Variation

SSV is a method to calculate if a mutation could be beneficial or not for an enzyme.

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SSV documentation

What is SSV?

SSV (Structural Signature Variation) is a method to propose mutations for enzymes used in industrial applications. **SSV** uses structural signatures to detect patterns, which can improve the activity of enzymes. A real and important application for SSV is the second-generation biofuel production.

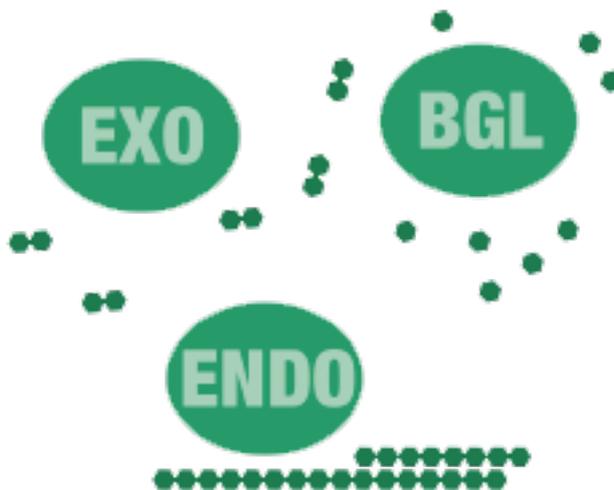


Case study: Second-Generation Biofuel problem

We used SSV as a method to propose mutations for β -glucosidase enzymes used in Second-Generation Biofuel production. SSV uses structural signatures to detect patterns, which can improve the activity of non-tolerant β -glucosidases, based on a manually curated database of glucose-tolerant β -glucosidases.

Second-Generation Biofuel is produced by residues obtained from the first generation biofuel production. They can be obtained from biomass, such as corn, algae, sugarcane, and so on. The production process is based on the extraction of fermentable sugars from cellulose.

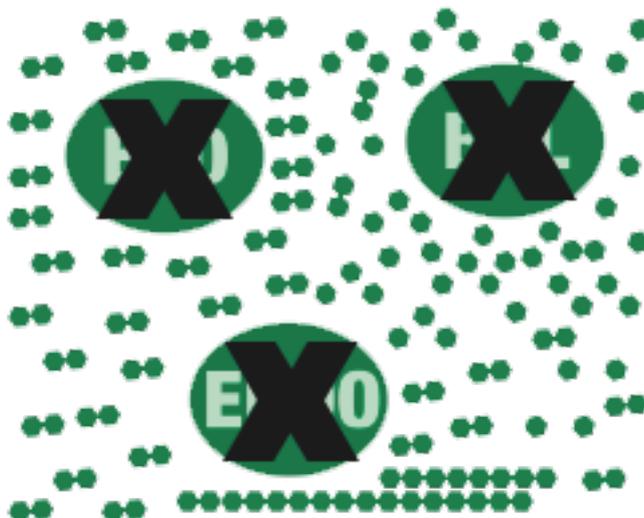
Cellulose is decomposed by the action of three enzymes: endoglucanases, exoglucanases, and β -glucosidases. Endoglucanases act first, cleaving cellulose in oligosaccharides of several lengths. Then, exoglucanases cleave the oligosaccharides in disaccharides, such as cellobiose. In the end, beta-glucosidases cleaves the cellobiose in two molecules of glucose, that will be used in the fermentation process to obtaining biofuel.



β -glucosidases are strongly inhibited by high glucose concentrations (same used for biofuel production).



In addition, it increases cellobiose concentration that inhibits endoglucanases and exoglucanases.

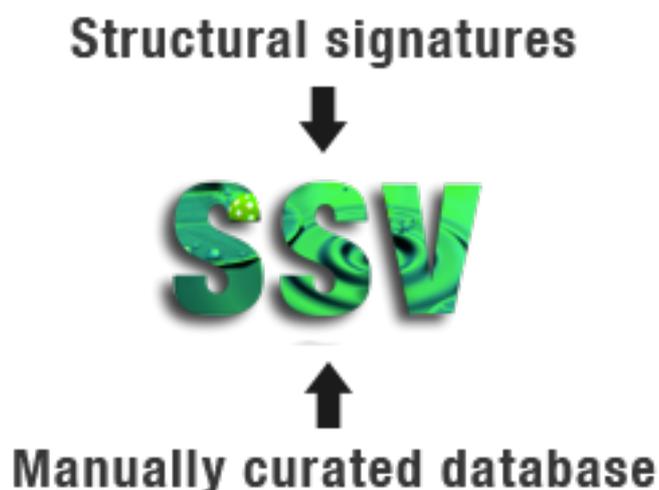


Glucose-tolerant β -glucosidases: β -glucosidases of high resistance to glucose inhibition, also called glucose-tolerants, can help to improve the biofuel production. Also, some mutations can turn non-tolerant in glucose-tolerant β -glucosidases.



How does SSV method work?

SSV method constructs structural signatures for wild and mutant proteins and compares the signature's variation with a manually curated database. In the case study, we used a database of glucose-tolerant β -glucosidases, called **Betagdb** (available at <http://bioinfo.dcc.ufmg.br/betagdb>).



Inputs

SSV requires three proteins:

1. Wild protein

2. Mutant protein
3. Template (reference)

SSV steps for the case study



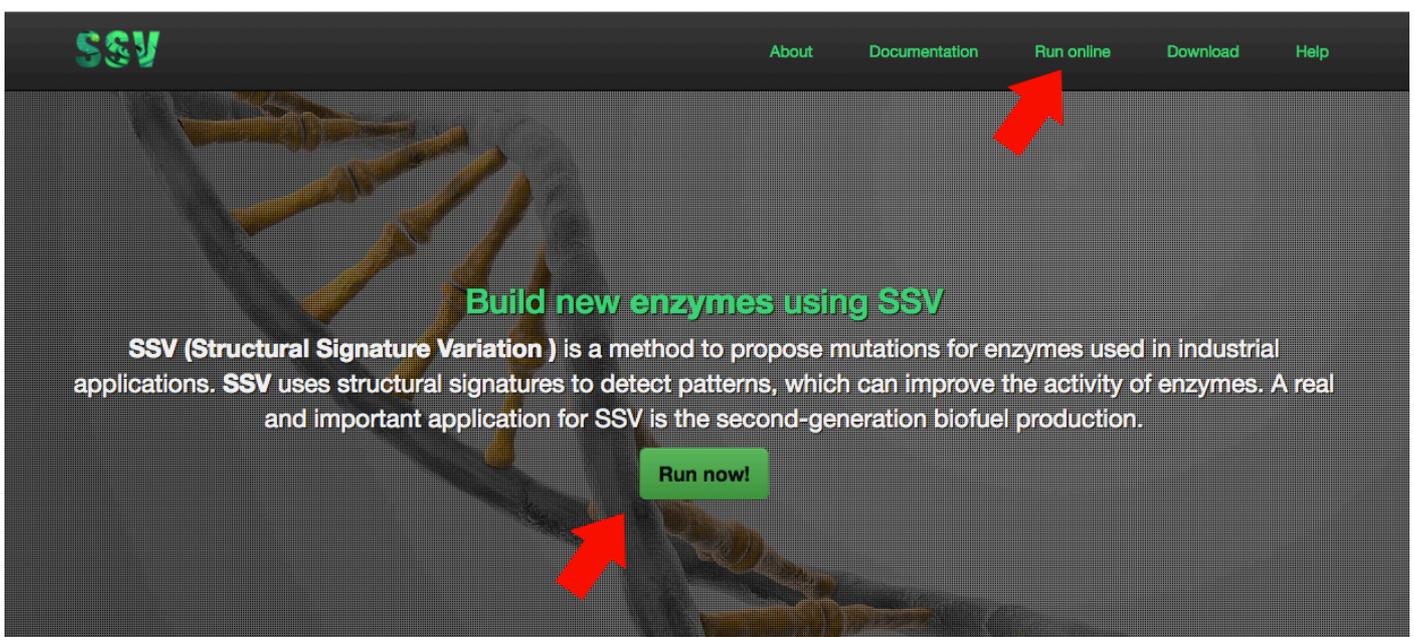
1. We constructed structural signatures using aCSM-ALL. We used cutoff ranging from 0Å until 10Å, and step distance of 0.1Å. However, we did not use the whole protein, only the region near to the active site. First, we performed the extraction of the catalytic pocket residues. This is not necessary for all projects run in SSV, however, if you use the whole protein, this increase the computational costs. The mutant and wild protein are aligned with 3VIK, the only beta-glucosidase in complex with cellobiose detected in the literature. The residues of the catalytic pocket of 3VIK are known: Q45, H148, W149, N192, S193, L195, T196, D199, M207, N253, I254, N255, Y273, N335, F336, Y337, T338, L340, W374, E402, W444, E451, W452, and F460. We performed alignments using MultiProt (<http://bioinfo3d.cs.tau.ac.il/MultiProt>);
2. We previously calculated the signature of 23 glucose-tolerant beta-glucosidases of betagdb. We calculated the distance among all glucose-tolerant beta-glucosidase and wild and mutant. The lowest values correspond to the templates;
3. We calculated the distance between wild and its template (ΔSSV_{wt}), and mutant and template (ΔSSV_{Mt}). The difference between both distances is the variation of the signature variations ($\Delta\Delta SSV$).

Running online

Running online is the best way to execute GTS. It is available online at: <<http://bioinfo.dcc.ufmg.br/gts>>.

Accessing the “Run online” panel

Click on “Run online” or in the “Run now!” Button.



The “Run online” panel requires some information to execute:

- **Project name:** define a name for your project. You can use any name (the system will create a unique ID for each project);
- **E-mail:** declare your e-mail (optional);
- **Mutation evaluated:** insert the point mutation or multiple mutations evaluated (optional);
- **Wild PDB:** input the wild PDB file;
- **Mutant PDB:** input the mutant PDB file;
- **Templates database:** input a set of proteins that it will be used as templates (send a zip file).

Run online

Project name:
Define a name for your project (required)

E-mail:
Receive the results in your e-mail (optional)

Mutations evaluated:
E.g. E96K (optional)

Wild PDB (required):
Escolher arquivo Nenhum arquivo selecionado

Mutant PDB (required):
Escolher arquivo Nenhum arquivo selecionado

Limit: 2MB. Please, send only one chain.

Templates database (required; ZIP format):
Escolher arquivo Nenhum arquivo selecionado

[Download sample dataset](#)

Submit

We made available a database with 27 mutations evaluated in the paper and the 23 proteins of betagdb (the templates for the case study). We can download the database and perform analysis using SSV online. We also made available a table with expected values and the values found.

SSV
Load Help

Download sample dataset [Download](#) ✕

id	File (Wild)	File (Mutant)	Mutation	$\Delta\Delta\text{GTS}$ expected	$\Delta\Delta\text{GTS}$ score
1	w1.pdb	m1.pdb	H228T	$\Delta\Delta\text{GTS} < 0$	-186.18
2	w2.pdb	m2.pdb	V174C/A404V/L441F	$\Delta\Delta\text{GTS} < 0$	-246.22
3	w3.pdb	m3.pdb	H184F	$\Delta\Delta\text{GTS} < 0$	100.37
4	w4.pdb	m4.pdb	P172L	$\Delta\Delta\text{GTS} < 0$	-6.29
5	w5.pdb	m5.pdb	P172L/F250A	$\Delta\Delta\text{GTS} < 0$	-6.29
6	w6.pdb	m6.pdb	L167W	$\Delta\Delta\text{GTS} < 0$	-602.80
7	w7.pdb	m7.pdb	L167W/P172L	$\Delta\Delta\text{GTS} < 0$	-615.46
8	w8.pdb	m8.pdb	L167W/P172L/P338F	$\Delta\Delta\text{GTS} < 0$	-615.46
9	w9.pdb	m9.pdb	V168Y	$\Delta\Delta\text{GTS} > 0$	330.56
10	w10.pdb	m10.pdb	F225S	$\Delta\Delta\text{GTS} > 0$	-365.07
11	w11.pdb	m11.pdb	Y308F	$\Delta\Delta\text{GTS} > 0$	34.19
12	w12.pdb	m12.pdb	Y308A	$\Delta\Delta\text{GTS} > 0$	-108.62
13	w13.pdb	m13.pdb	I207V	$\Delta\Delta\text{GTS} < 0$	-71.56
14	w14.pdb	m14.pdb	N218H	$\Delta\Delta\text{GTS} < 0$	-230.61
15	w15.pdb	m15.pdb	N273V	$\Delta\Delta\text{GTS} > 0$	-55.26
16	w16.pdb	m16.pdb	F252I	$\Delta\Delta\text{GTS} > 0$	86.70

Running an example (H228T)

Now, we will run the first example of that database. The mutation H228T, where detected for a non-tolerant beta-glucosidase and improved its glucose tolerance. For this reason, we expected a $\Delta\Delta\text{SSV}$ negative.

Download the dataset:

bioinfo.dcc.ufmg.br/ssv/#run

Download sample dataset

id	File (Wild)	File (Mutant)	Mutation	$\Delta\Delta\text{GTS}$ expected	$\Delta\Delta\text{GTS}$ score
1	w1.pdb	m1.pdb	H228T	$\Delta\Delta\text{GTS} < 0$	-186.18
2	w2.pdb	m2.pdb	V174C/A404V/L441F	$\Delta\Delta\text{GTS} < 0$	-246.22
3	w3.pdb	m3.pdb	H184F	$\Delta\Delta\text{GTS} < 0$	100.37
4	w4.pdb	m4.pdb	P172L	$\Delta\Delta\text{GTS} < 0$	-6.29
5	w5.pdb	m5.pdb	P172L/F250A	$\Delta\Delta\text{GTS} < 0$	-6.29
6	w6.pdb	m6.pdb	L167W	$\Delta\Delta\text{GTS} < 0$	-602.80
7	w7.pdb	m7.pdb	L167W/P172L	$\Delta\Delta\text{GTS} < 0$	-615.46
8	w8.pdb	m8.pdb	L167W/P172L/P338F	$\Delta\Delta\text{GTS} < 0$	-615.46
9	w9.pdb	m9.pdb	V168Y	$\Delta\Delta\text{GTS} > 0$	330.56
10	w10.pdb	m10.pdb	F225S	$\Delta\Delta\text{GTS} > 0$	-365.07
11	w11.pdb	m11.pdb	Y308F	$\Delta\Delta\text{GTS} > 0$	34.19
12	w12.pdb	m12.pdb	Y308A	$\Delta\Delta\text{GTS} > 0$	-108.62
13	w13.pdb	m13.pdb	I207V	$\Delta\Delta\text{GTS} < 0$	-71.56
14	w14.pdb	m14.pdb	N218H	$\Delta\Delta\text{GTS} < 0$	-230.61
15	w15.pdb	m15.pdb	N273V	$\Delta\Delta\text{GTS} > 0$	-55.26

dataset.zip

Extract the files:

dataset

Nome	Data de Modificação	Tamanho	Tipo
m1.pdb	8 de ago de 2016 16:01	16 KB	Docum...t 2.app
m2.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m3.pdb	8 de ago de 2016 16:02	17 KB	Docum...t 2.app
m4.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m5.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m6.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m7.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m8.pdb	8 de ago de 2016 16:02	18 KB	Docum...t 2.app
m9.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m10.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m11.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m12.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m13.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m14.pdb	8 de ago de 2016 16:02	17 KB	Docum...t 2.app
m15.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m16.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m17.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m18.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m19.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m20.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m21.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m22.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m23.pdb	8 de ago de 2016 16:02	18 KB	Docum...t 2.app
m24.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
m25.pdb	8 de ago de 2016 16:01	17 KB	Docum...t 2.app
m26.pdb	8 de ago de 2016 16:02	17 KB	Docum...t 2.app
m27.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app
templates.zip	Ontem 16:00	93 KB	ZIP
w1.pdb	8 de ago de 2016 16:02	17 KB	Docum...t 2.app
w2.pdb	8 de ago de 2016 16:02	17 KB	Docum...t 2.app
w3.pdb	8 de ago de 2016 16:02	17 KB	Docum...t 2.app
w4.pdb	8 de ago de 2016 16:01	18 KB	Docum...t 2.app

Run SSV using w1.pdb (wild), m1.pdb (mutant), and templates.zip (23 templates):



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Run online

Nenhum arquivo selecionado

Project name:

E-mail:

Mutations evaluated:

Wild PDB (required):
 | w1.pdb

Mutant PDB (required):
 | m1.pdb
Limit: 2MB. Please, send only one chain.

Templates database (required; ZIP format):
 | templates.zip

[Download sample dataset](#)

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After submitting the data, SSV online will process your requisition. A unique ID will be created for your project. When the process to finish, you can click at the link.



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Your project was created. You can access it at the link: [SSV48B6462](#)

While SSV runs your project, you will receive this message:



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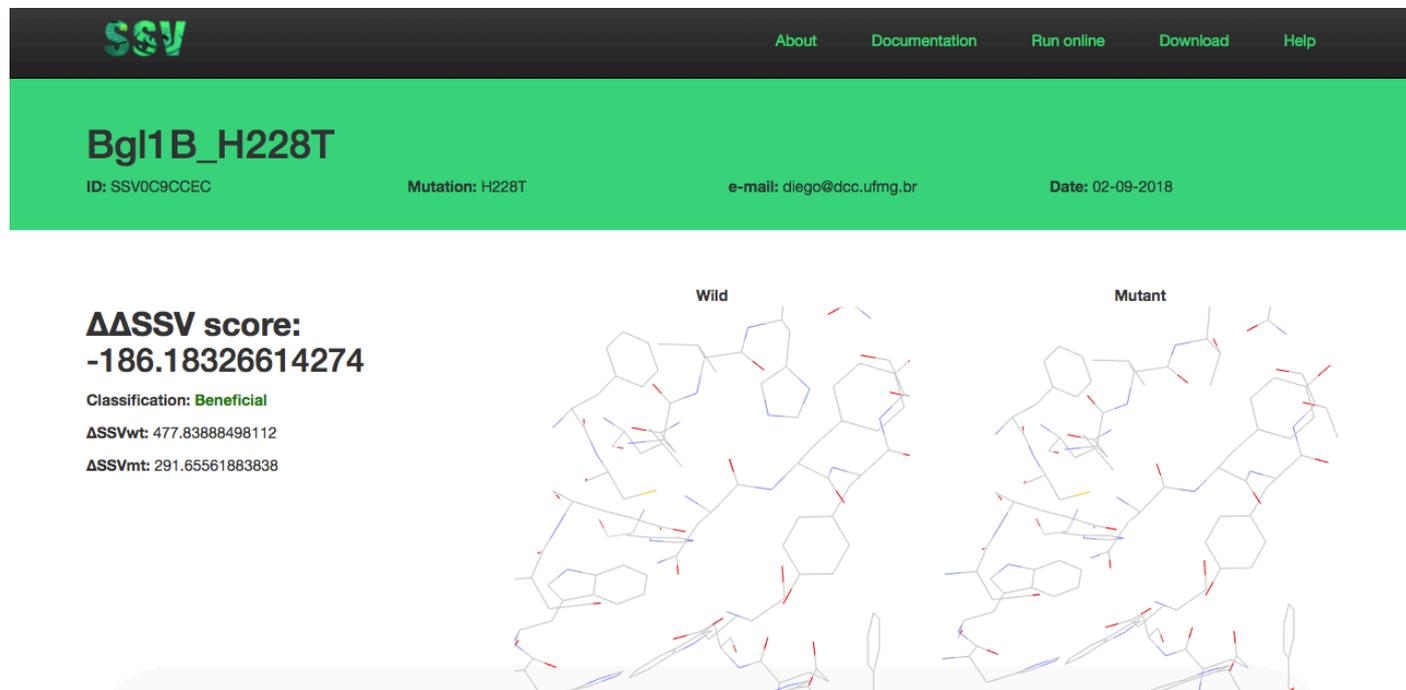
Your project is processing. Try again later. You can access it at the link: [SSV0C9CCEC](#)

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In the end, you will be redirected to the individual page of the project. This page is identified by the unique ID, described under the project name (in the green sector).

Below the green sector, there are three important sectors:

1. The results of the $\Delta\Delta\text{SSV}$ calculation;
2. Wild visualization;
3. Mutant visualization.



In the main panel, it is shown:

- $\Delta\Delta\text{SSV}$ score;
- Classification;
- $\Delta\text{SSV}_{\text{wt}}$;
- $\Delta\text{SSV}_{\text{Mt}}$.

Run by scripts

We made available the scripts to run SSV. However, we cannot have sure that you will be able to reproduce the software environment. If you have problems, run the online version.

We strongly recommend running SSV online.

Requirements

- O.S. Linux 64bit (recommended Ubuntu 16.04)
- Python (version 2)
- Library Numpy
- Perl
- aCSM
- MultiProt

Scripts

Script	Input	Output
extractSite.py	PDBs files inserted into the folder "pdb".	PDBs files with the catalytic pocket extracted.
aCSM.pl	List with the addresses of PDBs files with the catalytic pocket extracted.	CSV file with the total of atoms in the cutoff distances.
dgts3.py	CSV file obtained from aCSM. The first 23 lines must represent the beta-glucosidases from betagdb in alphabetical order.	$\Delta\Delta$ SSV and templates for wild and mutant save in the file "result.txt".